



## Massive Cortical Reorganization After Sensory Deafferentation in Adult Macaques

Tim P. Pons; Preston E. Garraghty; Alexander K. Ommaya; Jon H. Kaas; Edward Taub; Mortimer Mishkin

*Science*, New Series, Vol. 252, No. 5014. (Jun. 28, 1991), pp. 1857-1860.

Stable URL:

<http://links.jstor.org/sici?sici=0036-8075%2819910628%293%3A252%3A5014%3C1857%3AMCRASD%3E2.0.CO%3B2-5>

*Science* is currently published by American Association for the Advancement of Science.

---

Your use of the JSTOR archive indicates your acceptance of JSTOR's Terms and Conditions of Use, available at <http://www.jstor.org/about/terms.html>. JSTOR's Terms and Conditions of Use provides, in part, that unless you have obtained prior permission, you may not download an entire issue of a journal or multiple copies of articles, and you may use content in the JSTOR archive only for your personal, non-commercial use.

Please contact the publisher regarding any further use of this work. Publisher contact information may be obtained at <http://www.jstor.org/journals/aaas.html>.

Each copy of any part of a JSTOR transmission must contain the same copyright notice that appears on the screen or printed page of such transmission.

---

JSTOR is an independent not-for-profit organization dedicated to and preserving a digital archive of scholarly journals. For more information regarding JSTOR, please contact [support@jstor.org](mailto:support@jstor.org).

- possible significance of temporal structure and correlated firing in the visual cortex [C. M. Gray and W. Singer, *Proc. Natl. Acad. Sci. U.S.A.* **86**, 1698 (1989)]. All these observations, however, involve measurements of average properties of spike trains over repeated presentations of a stimulus. Although these experiments are suggestive of coding strategies that different systems might use, none directly answers the question of how the organism could extract this coded information from single spike trains in real time.
5. W. Bialek, in *1989 Lectures in Complex Systems, SFI Studies in the Sciences of Complexity*, E. Jen, Ed. (Addison-Wesley, Reading, MA, 1990), vol. 2, pp. 513-595.
  6. The most general of such approaches are the white-noise methods, such as reverse correlation [P. Marmarelis and V. Marmarelis, *Analysis of Physiological Systems. The White Noise Approach* (Plenum, New York, 1978)]. Although these methods do not rely on repeated presentation of identical stimuli, they nonetheless yield models that ideally predict the firing rate in response to arbitrarily chosen stimuli. Reverse correlation thus suffers the same limitations as other rate-based approaches.
  7. Other authors have realized the importance of approaching neural coding from the point of view of the organism. In early work, R. Fitzhugh [*J. Gen. Physiol.* **41**, 675 (1958)] discusses real-time decision making. As far as we know, P. I. M. Johannesma [in G. Székely, E. Lábos, S. Damjanovich, Eds., *Adv. Physiol. Sci.* **30**, 103 (1981); Akadémiai Kiadó, Budapest] comes closest to our approach.
  8. F. Rieke, W. Yamada, K. Moortgat, E. R. Lewis, W. Bialek, in *Auditory Physiology and Perception: Proceedings of the 1991 European Hearing Workshop*, in press.
  9. W. Bialek and A. Zee, *J. Stat. Phys.* **59**, 103 (1990).
  10. Linear decoding might work for a trivial reason. Many sensory neurons have a regime in which the firing rate varies linearly with stimulus amplitude. This is an example of linear encoding and linear decoding might work as well in this regime. However, Bialek and Zee (9) predict that linear decoding is possible beyond the regime of linear input-output relations. The cell we studied in this work has a linear input-output regime, but the stimuli we used drove the cell outside this limit into saturation. See, for example, H. Eckert [*J. Comp. Physiol. A* **135**, 29, (1980)], who finds nonmonotonic rate-velocity profiles in the range of stimuli used here.
  11. To ensure that the reconstruction process could be implemented in real time, we required that the filters be causal, for example,  $F_1(\tau < 0) = 0$ . We calculated the minimum  $\chi^2$  causal filters in two ways: (i) The best filter is first calculated without the causality constraint. An explicit formula can be written for this filter in terms of the spike trains and the stimulus

$$F_1(\tau) = \int \frac{d\omega}{2\pi} e^{-i\omega\tau} \frac{\langle \tilde{s}(\omega) \sum_j e^{-i\omega t_j} \rangle}{\langle \sum_{i,j} e^{i\omega(t_i - t_j)} \rangle} \quad (3)$$

- where  $\tilde{s}(\omega) = \langle d\tau \rightarrow \tilde{s}(\omega) = \int d\tau \rangle$ . The averages (inside triangular brackets) are over the ensemble of stimuli  $s(\tau)$  used in the experiment. This filter can be shifted by a delay  $\tau_{\text{delay}}$  and causality can be imposed by setting the shifted filter to zero at negative times. (ii)  $\chi^2$  can be minimized with respect to purely causal functions by expansion of the filters  $\{F_n\}$  in a complete set of functions that vanish at negative times. In this method, a delay time must be explicitly introduced that measures the lag between the true stimulus and the reconstruction, so  $\chi^2(\tau_{\text{delay}}) = \int dt |s(t - \tau_{\text{delay}}) - s_{\text{est}}(t)|^2$  is minimized. These two methods together ensure that the optimal causal filters are calculated.
12. D. Warland, M. A. Landolfa, J. P. Miller, W. Bialek, in *Analysis and Modeling of Neural Systems*, F. Eckman, Ed. (Kluwer, Norwell, MA, in press).
  13. K. Hausen, in *Photoreception and Vision in Invertebrates*, M. Ali, Ed. (Plenum, New York, 1984).
  14. M. Land and T. Collett, *J. Comp. Physiol.* **89**, 331 (1974).
  15. R. R. de Ruyter van Steveninck, *Real-Time Performance of a Movement-Sensitive Neuron in the Blowfly*

*Visual System* (Rijksuniversiteit Groningen, Groningen, the Netherlands, 1986).

16. C. Shannon, *Bell Syst. Tech. Jour.* **27**, 379 (1948).
17. G. Westheimer, *Invest. Ophthalmol.* **14**, 570 (1975).
18. W. Reichardt, in *Principles of Sensory Communication*, W. Rosenblith, Ed. (Wiley, New York, 1961).
19. C. Koch and I. Segev, Eds., *Methods in Neuronal Modeling* (MIT Press, Cambridge, MA, 1989).
20. C. Koch, T. Poggio, V. Torre, *Proc. Natl. Acad. Sci. U.S.A.*, **80**, 2799 (1983).
21. We thank W. J. Bruno, M. Crair, W. Gerstner, L. Kruglyak, J. P. Miller, W. G. Owen, A. Zee, and G. Zweig for helpful discussions and D. A. Baylor, D. Glaser, and M. Meister for thoughtful comments on the manuscript. Preliminary work on simulations of model neurons was done with L. Kruglyak, the results on the cricket cercal system

were obtained in collaboration with M. A. Landolfa and J. P. Miller, and the experiments on the bullfrog auditory system were done in collaboration with W. Yamada and E. R. Lewis. Work at Berkeley was supported in part by the NSF through a Presidential Young Investigator Award to W.B., supplemented by funds from Cray Research, Sun Microsystems, and the NEC Research Institute, and through a Graduate Fellowship to F.R. D.W. was supported in part by the Systems and Integrative Biology Training Program of the NIH. Initial work was supported by the Netherlands Organization for Pure Scientific Research (ZWO). Research also performed at NEC Research Institute by W.B. and F.R.

2 November 1990; accepted 10 May 1991

## Massive Cortical Reorganization After Sensory Deafferentation in Adult Macaques

TIM P. PONS,\* PRESTON E. GARRAGHTY, ALEXANDER K. OMMAYA, JON H. KAAS, EDWARD TAUB, MORTIMER MISHKIN

After limited sensory deafferentations in adult primates, somatosensory cortical maps reorganize over a distance of 1 to 2 millimeters mediolaterally, that is, in the dimension along which different body parts are represented. This amount of reorganization was considered to be an upper limit imposed by the size of the projection zones of individual thalamocortical axons, which typically also extend a mediolateral distance of 1 to 2 millimeters. However, after extensive long-term deafferentations in adult primates, changes in cortical maps were found to be an order of magnitude greater than those previously described. These results show the need for a reevaluation of both the upper limit of cortical reorganization in adult primates and the mechanisms responsible for it.

MERZENICH AND HIS COLLEAGUES demonstrated that primary cortical sensory maps in adult animals, like those in infant animals, are capable of reorganization after various peripheral sensory perturbations (1, 2). Yet, compared to the massive functional changes that have been found in neonates, in which entire cortical maps may be reorganized (3), the changes reported in adults have been relatively small, with an upper limit of 1 to 2 mm along the cortical surface (1, 2, 4). Although the finding of any plasticity in primary sensory maps of adult animals was unexpected, the limited extent of the changes suggested they were confined to the projection zones of single thalamocortical axons (1, 2). Both the limits of reorganization and the mechanisms responsible must now be reconsidered because of new evidence in adult macaques showing reorganization in the cortex at least an order of

magnitude greater than that reported previously.

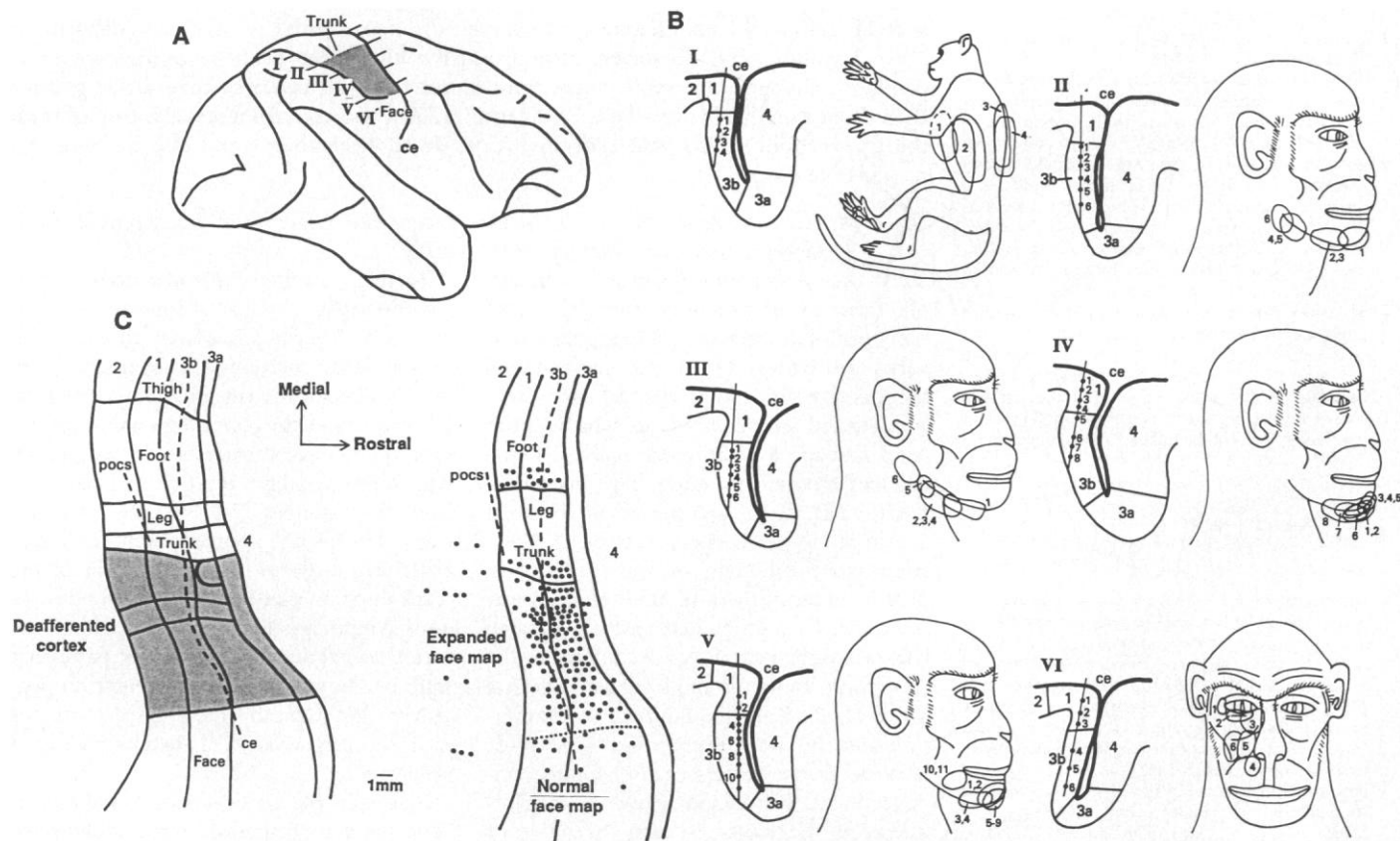
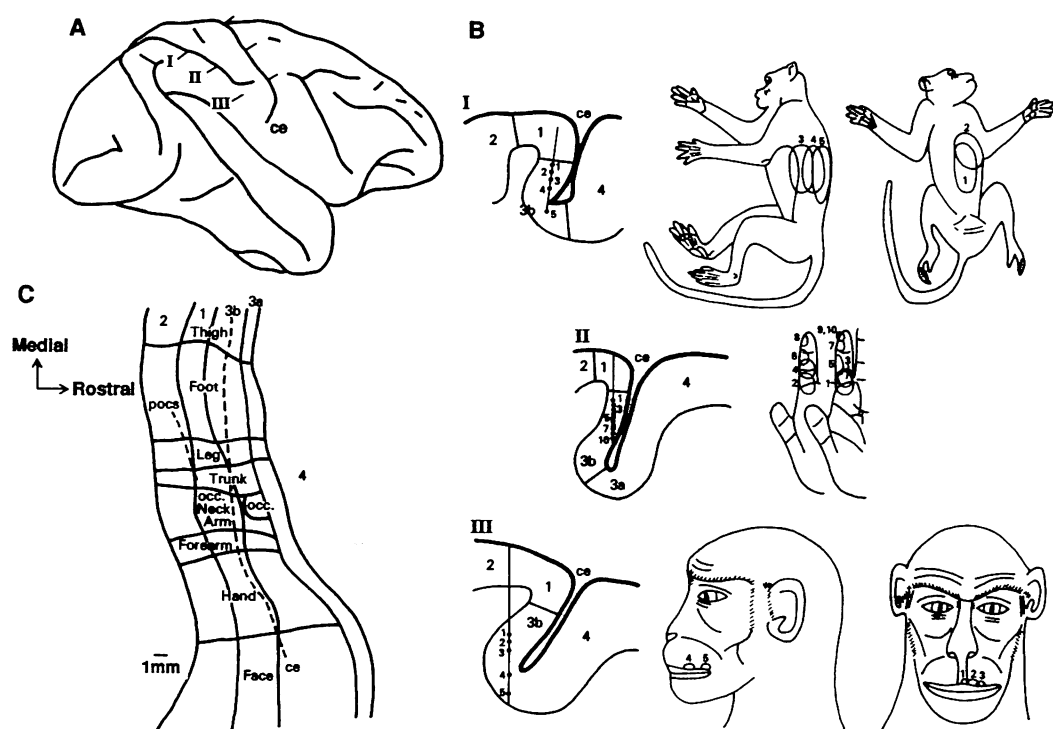
Tactually elicited neuronal activity was recorded in area SI (5) of four cynomolgus monkeys (*Macaca fascicularis*) that had received deafferentations of an upper limb, three unilateral and one bilateral, more than 12 years before the recording session (6). All procedures were carried out in accordance with NIH guidelines on the care and use of laboratory animals (7). Electrode penetrations were placed approximately 0.75 mm apart across the mediolateral extent of the cortical region that had been deprived of its normal input and less densely in parts of the cortex containing maps of body parts that were unaffected by the deafferentation procedure. We typically recorded activity for each 300- $\mu\text{m}$  advance of the electrode in a penetration.

Normally the cortical representations of body parts are organized into highly topographic maps (8, 9) (Fig. 1). In macaques, the upper limb representation in SI is always bordered by the representation of the trunk medially and the face laterally (10). In the region of the border of the face and hand representations, which is located opposite the tip of the intraparietal sulcus (8), the face map contains the representation of the chin

T. P. Pons, A. K. Ommaya, M. Mishkin, Laboratory of Neuropsychology, National Institute of Mental Health, Building 9, Room 1N107, Bethesda, MD 20892.  
P. E. Garraghty and J. H. Kaas, Department of Psychology, Vanderbilt University, Nashville, TN 37240.  
E. Taub, Department of Psychology, University of Alabama at Birmingham, Birmingham, AL 35294.

\*To whom correspondence should be addressed.

**Fig. 1.** (A) Lateral brain view indicating location of the three parasagittal sections (I through III) illustrated on the right. (B) Three parasagittal sections through SI of a normal animal showing electrode tracks (vertical lines) and recording sites (dots). Numbered recording sites (not all are identified by a numeral because of a lack of space) correspond to numbered receptive fields on the body part shown to the right of the parasagittal section. As recording sites traverse the caudal bank of the central sulcus through area 3b, receptive fields on both the trunk (I) and face (III) shift laterally away from ventral midline, while those on the digits (II) shift distally. (C) Flattened map of SI showing normal somatotopy. Body part borders are marked by horizontal lines, areal borders by vertical lines, and central and post-central sulci by dashed vertical lines. The following abbreviations are used: ce, central sulcus, pocs, postcentral sulcus, and occ., occiput. Figure modified from (8).



**Fig. 2.** (A) Lateral brain view showing the portion of the postcentral cortex that was deprived of its normal inputs by the deafferentation procedure ("deafferented zone" marked by shading) and the locations of the six parasagittal sections (I through VI) illustrated on the left. (B) Sections I and VI show receptive field data from the cortex medial and lateral, respectively, to the deafferented zone. The normal receptive field progression from ventral midline to lateral body parts across the trunk and face was encountered as the electrode traversed the caudal bank of the central sulcus through area 3b (compare with Fig. 1). Sections II through V show the portion of the face

represented across the deafferented zone. In section II, located immediately adjacent to the trunk representation, recording sites were still responsive to stimulation of the face. Also, the normal sequence of receptive fields from the ventral midline (chin) to the lateral parts of face (lower jaw) was apparent as recording sites traversed area 3b, as was the mirror reversal of this sequence in area 1 (sections IV through VI). (C) Two flattened maps of SI, the first showing the deafferented zone (marked by shading), and the second the recording site density in the animal illustrated (CM3). Other conventions as in Fig. 1.

and lower jaw and the hand map contains the representation of the thumb. The entire upper-limb representation extends lateromedially for 10 to 14 mm, from the lateral tip of the intraparietal sulcus to the lateral tip of the postcentral sulcus, where the trunk representation is normally found.

The area of the cortex deprived of its normal input by the deafferentation procedure, which we refer to as the deafferented zone, included the SI maps of the fingers, palm, remaining upper limb, neck, and occiput (Fig. 2) (8). Our recordings unexpectedly revealed that this entire zone responded to stimulation of the face. In the animal illustrated in Fig. 2, we were able to obtain vigorous neuronal responses to light stimulation of the face in 124 recording sites distributed throughout the deafferented zone. Furthermore, none of the sites we tested was unresponsive.

Virtually identical findings were obtained in the three other animals. All 320 sites tested in the deafferented zone in the four animals were activated by face stimulation. An additional 90 and 51 recording sites located lateral and medial, respectively, to the deafferented zone revealed the expected normal topography of face and trunk (8, 10). Thus, in all cases, the medial border of the expanded face representation abutted the normal representation of the trunk. There was no apparent elevation of response thresholds at any of the recording sites across the new face map as compared to those across the normal face map; in both, a slight deflection of facial hairs was sufficient to obtain a vigorous neuronal response.

Not all of the face, however, was represented in the reorganized region; rather, stimulation of only a relatively small portion of the face, from the chin to the lower jaw, was found to activate neurons in this zone. At the same time, the pattern of reorganization in this new part of the face map was not random but highly systematic. As in normal face maps (Fig. 1), the midline of the face, in this case the chin, was represented caudally in area 3b (that is, near the border of areas 1 and 3b), whereas progressively more lateral parts of the face, in this case the lateral parts of the lower jaw, were represented in progressively more rostral parts of area 3b (that is, toward the border of areas 3b and 3a). Normally, the representation of the chin and lower jaw is located immediately adjacent to the hand representation. Consequently, it appeared as though each point on the normal face map along the original border of the hand and face representations had been stretched medially into a line approximately 10 to 14 mm long, the length of the deafferented zone. This resulted in the apparent stretching of the entire chin and lower jaw

map [at least in areas 3b and 1 (5)] onto a cortical sheet 10 to 14 mm long, until the expanded face representation met the normal trunk map (11). These findings extend the previously proposed upper limit (4, 12) for reorganization in adult primates by an order of magnitude and leave open the possibility that the limit is even greater.

What mechanisms could account for such massive cortical reorganization in mature animals? In earlier studies on the effects of peripheral deafferentations in adult primates, the deafferentations were relatively restricted, involving small parts of the hand (1, 2) or visual field (13), and the deafferented zone came to represent the sensory surfaces mapped along the zone's lateral and medial edges, with each of these two representations expanding toward the deafferented zone's center. Furthermore, the occupation of the deafferented zone by these new inputs was often incomplete, with small islands of tissue remaining unresponsive to stimulation of any body part (1, 2, 4). Because of those features and the spatial limit of reorganization, which was generally in the range of 1 to 2 mm, it was reasonable to relate the filling in of the map to the mediolateral arborization of single thalamocortical axons (1, 2), which is also in the range of 1 to 2 mm (14). Because of the spatial extent of such arborization, neurons at a given cortical site could receive overlapping thalamic projections from two populations of axons, one representing a dominant skin region and the other an adjacent, non-dominant skin region; if so, then loss of the former would allow neuronal activation by the latter, either immediately or after a delay. Although such a mechanism may suffice for the limited changes described previously, it is insufficient to account for the extensive reorganization reported here.

An alternative possibility is that preexisting inputs from face representations in cortical areas outside SI came to activate the deafferented zone. Such a possibility seems remote, however, because all connections of these areas with SI are between somatotopically matched representations (15), a circumstance that should impose the same constraints on reorganization as the somatotopically matched thalamocortical projections. If the reorganization we found took place exclusively at the cortical level (1, 2, 4), then the only alternative to the immediate or delayed unmasking (16) of preexisting thalamocortical or corticocortical projections would be the sprouting of new projections across the deafferented zone. Yet there is no evidence to date of even limited sprouting of sensory terminals in the neocortex after peripheral nervous system injury in adult mammals.

These considerations lead us to propose that much of the functional reorganization we observed was a reflection of changes that had taken place subcortically and were then simply relayed to the cortex (17). Body part maps are represented within a much smaller neural space in the brain stem than in the thalamus, and in the thalamus than in the cortex, reflecting the extensive divergence that occurs along pathways connecting the brainstem, thalamus, and cortex (18); as a result, reorganization over a relatively small distance at the brain stem or thalamic levels would be reflected as much larger changes at the cortical level (19). Thus, if projections to or from brain stem nuclei representing the face were to have synapsed onto all or most of the brain stem or thalamic cells that had previously represented the upper limb, then the entire upper limb representation in the cortex would likewise have come to represent the face. Furthermore, axonal sprouting after deafferentation has been reported to occur in the spinal cord (20), making it more plausible that such changes could also be taking place at higher subcortical stations.

Our finding of extensive reorganization after peripheral deafferentation raises many additional questions. For example, why was the deafferented zone not occupied by an expanded trunk as well as by an expanded face representation (21)? Did the expanded face representation mediate tactile perception, and could it serve as a substitute for the normal face representation? Was the neural activity in the expanded representation relayed to higher order cortical and subcortical stations? Answers to such questions about mechanism and function could lead to harnessing the immense reorganizational capability of the adult nervous system for therapeutic purposes (22).

#### REFERENCES AND NOTES

1. M. M. Merzenich *et al.*, *Neuroscience* **10**, 639 (1983).
2. ———, *ibid.* **8**, 33 (1983); M. M. Merzenich and J. H. Kaas, *Trends Neurosci.* **5**, 434 (1982); J. H. Kaas, M. M. Merzenich, H. P. Killackey, *Annu. Rev. Neurosci.* **6**, 325 (1983).
3. S. M. Sherman and P. D. Spear, *Physiol. Rev.* **62**, 738 (1982); A. W. Roe, S. L. Pallas, J. O. Hahn, M. Sur, *Science* **250**, 818 (1990).
4. M. M. Merzenich *et al.*, in *Neurobiology of Neocortex*, P. Rakic and W. Singer, Eds. (Wiley, New York, 1988), pp. 41–67.
5. SI in macaques includes four cytoarchitectonic areas (3a, 3b, 1, and 2). Area 3b of primates is the homolog of SI in lower mammals [J. H. Kaas, *Physiol. Rev.* **63**, 206 (1983)]. We concentrated our recordings on area 3b and to a lesser extent area 1, because more is known about their normal organization than about that of areas 3a and 2. Areas 3b and 1 combined extend rostrocaudally approximately 8 to 10 mm.
6. The deafferentations, performed when the animals were 3 to 4 years old, involved transection of the dorsal roots of spinal segments C2 to T4 at the point where they entered the cord. For details of surgery see E. Taub, in *Behavioral Psychology in Rehabilitation*

tion *Medicine: Clinical Applications*, L. P. Ince, Ed. (Williams & Wilkins, New York, 1980), pp. 371–401.

7. Surgical and recording procedures are described in detail elsewhere (8). Animals were anesthetized with ketamine hydrochloride (25 to 50 mg per kilogram of body weight) before intubation, after which they were given a 1.5% mixture of isoflurane gas with O<sub>2</sub>. After placement in a stereotaxic frame a craniotomy was performed over the dorsolateral cortex contralateral to the deafferentation. Single- and multiunit neuronal activity was recorded with tungsten microelectrodes (1.0 to 1.5 megohms, measured at 1 kHz). We used a surgical microscope to sight the placement of the electrode on the surface of the cortex, and the electrode was then advanced by a hydraulic microdrive down the caudal bank of the central sulcus. Body parts were stimulated by gentle brushing with a cotton swab or camel hair brush.
8. R. J. Nelson, M. Sur, D. J. Felleman, J. H. Kaas, *J. Comp. Neurol.* **192**, 611 (1980); T. P. Pons, P. E. Garraghty, C. G. Cusick, J. H. Kaas, *ibid.* **241**, 445 (1985); T. P. Pons, J. T. Wall, P. E. Garraghty, C. G. Cusick, J. H. Kaas, *Somatosens. Res.* **4**, 309 (1987).
9. C. N. Woolsey, W. H. Marshall, P. Bard, *Bull. Johns Hopkins Hosp.* **70**, 339 (1942); G. Werner and B. L. Whitsel, *J. Neurophysiol.* **31**, 856 (1968); J. H. Kaas, R. J. Nelson, M. Sur, C. S. Lin, M. M. Merzenich, *Science* **204**, 521 (1979).
10. Although there is debate about the somatotopic

organization in the rostrocaudal dimension of SI, there is universal agreement about the topographic organization in the mediolateral dimension.

11. There was no obvious decrease in receptive field size for recording sites in the reorganized cortex; thus, the rule relating receptive field size to cortical magnification did not apply [M. Sur, M. M. Merzenich, J. H. Kaas, *J. Neurophysiol.* **44**, 295 (1980)].
12. Corroboration that this limit for reorganization can be exceeded is provided by P. E. Garraghty and J. H. Kaas, *Proc. Natl. Acad. Sci. U.S.A.*, in press.
13. S. J. Heinen and A. A. Skavenski, *Invest. Ophthalmol. Vis. Sci. Suppl.* **29**, 23 (1988); J. H. Kaas et al., *Science* **248**, 229 (1990).
14. P. E. Garraghty, T. P. Pons, M. Sur, J. H. Kaas, *Somatosens. Mot. Res.* **6**, 401 (1989); M. Conley and E. G. Jones, *Soc. Neurosci. Abstr.* **10**, 495 (1984); P. E. Garraghty and M. Sur, *J. Comp. Neurol.* **294**, 583 (1990). More extensive arborizations have been seen (E. G. Jones, personal communication), but not in a topographic pattern that would account for our results.
15. E. G. Jones, J. D. Coulter, S. H. C. Hendry, *J. Comp. Neurol.* **181**, 297 (1978); T. P. Pons and J. H. Kaas, *ibid.* **248**, 313 (1986); D. P. Friedman, E. A. Murray, J. B. O'Neill, *ibid.* **252**, 323 (1986).
16. R. W. Rhoades, G. R. Belford, H. P. Killackey, *J. Neurophysiol.* **57**, 1577 (1987).
17. The timing of these changes is unknown, but they are unlikely to have occurred soon after surgery, because even the limited "filling in" seen in earlier

studies was still incomplete several months postoperatively.

18. J. H. Kaas and T. P. Pons, in *Comparative Primate Biology, Neurosciences*, H. D. Steklis and J. Erwin, Eds. (Liss, New York, 1988), pp. 421–468; V. B. Mountcastle, *Handbook of Physiology, The Nervous System*, (American Physiological Association, Bethesda, MD 1984), vol. 3, pp. 789–878.
19. H. P. Killackey, *J. Cogn. Neurosci.* **1**, 3 (1989).
20. M. E. Goldberger and M. Murray, *Physiologic Basis for Functional Recovery in Neurological Disease*, S. G. Waxman, Ed. (Raven, New York, 1988), pp. 361–392; A. El-Bohy, S. E. Kapaıda, C. C. Lamotte, *Soc. Neurosci. Abstr.* **16**, 1162 (1990).
21. Both the extent and the selectivity of the expansion of the face representation was unexpected, because only small-scale changes had been seen at the SI face-hand border [figure 14, p. 656, in (1)].
22. For example, inputs normally processed by regions of the sensory cortex damaged by stroke might be rechanneled for processing by undamaged regions of sensory cortex.
23. We thank R. E. Burke, M. E. Goldberger, E. G. Jones, P. L. Strick, and W. D. Willis, Jr., for their contributions to the design of the research, J. L. Blanchard, H. B. Cavirac, and M. S. Raterec for their help and support in the conduct of the research and J. Brady, P. J. Gerone, F. A. King, and W. F. Raub for helping to make the research possible.

6 March 1991; accepted 26 April 1991

## 1991

### AAAS Philip Hauge Abelson Prize

The AAAS Philip Hauge Abelson Prize of \$2,500, established by the AAAS Board of Directors in 1985, is awarded annually either to:

- a **public servant**, in recognition of sustained exceptional contributions to advancing science, or
- a **scientist** whose career has been distinguished both for scientific achievement and for other notable services to the scientific community.

AAAS members are invited to submit nominations now for the 1991 prize, to be awarded at the 1992 Annual Meeting in Chicago, IL. Each nomination must be seconded by at least two other AAAS members.

Nominations should be typed and should include the following information: nominee's typed name, institutional affiliation and title, address, and brief biographical resume (please do not send lengthy publications lists); statement of justification for nomination; and names, identification, and signatures of the three or more AAAS member sponsors.

Nominations should be submitted to Mark S. Frankel, AAAS Directorate for Science and Policy Programs, 1333 H Street, N.W., Washington, D.C. 20005, for receipt by **1 August 1990**.

The winner will be selected by a seven-member selection panel. The award recipient is reimbursed for travel and hotel expenses incurred in attending the award presentation.